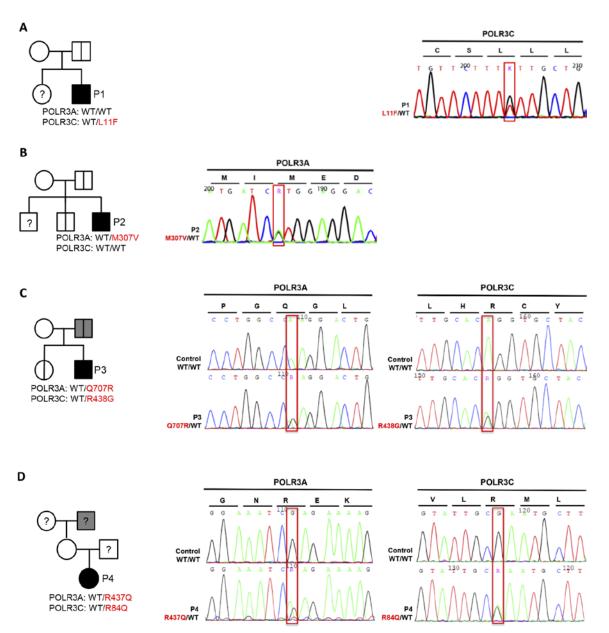
Supplemental material



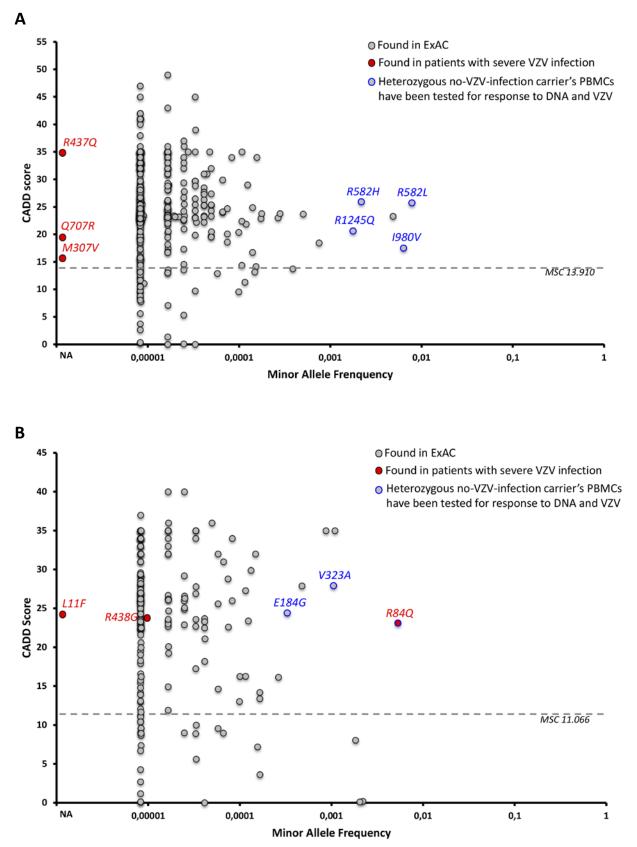
Supplemental Figure 1. Sanger sequencing of *POLR3A* **and** *POLR3C***.** DNA sequences of the regions in the POLR3A and C genes carrying the mutations.

POLR3A

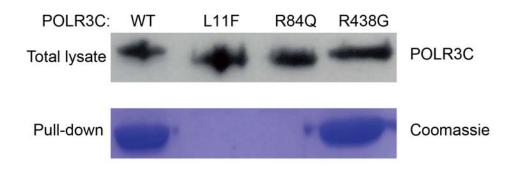
P2 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovis aries Felis catus	293 322 IKKHRISGAKTQMI EDWDFLQLQCALYIN IKKHRISGAKTQMINEDWDFLQLQCALYIN	P1 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovis aries Felis catus	1 30 MTQAEIKLCS LLQEHFGEIVEKIGVHLIR MTQAEIKLCS LLQEHFGEIVEKIGVHLIR
P3 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovis aries Felis catus	693 722 NRKPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY NRGPSIGIGDVTPCGGLIKAKYELINAGKY	P3 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovis aries Felis catus	70 99 RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTAKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY RGVVEYEAQCSRVLPMLRYPRYIYTKTLY
P4 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovis aries Felis catus	423 452 QRHTQMK RFLKYGN EKMAQELKYGDIVER QRHTQMK RFLKYGN EKMAQELKYGDIVER QRHTQMK RFLKYGN EKMAQELKYGDIVER QRHTQMK RFLKYGN EKMAQELKYGDIVER QRHTQMK RFLKYGN EKMAQELKFGDIVER QRHTQMK RFLKYGN EKMAQELKFGDIVER QRHMMKK RFLKYGN EKMAQELKFGDIVER QRHMMK RFLKYGN EKMAQELKYGDIVER QRHMQMK RFLKYGN EKMAQELKYGDIVER QRHTQMK RFLKYGN EKMAQELKYGDIVER	P4 Homo sapiens Macaca mulatta Pan troglodytes Bos taurus Sus scrofa Equus caballus Oryctolagus cuniculus Rattus norvegicus Mus musculus Otolemur garnettii Monodelphis domestica Papio anubis Callithrix jacchus Canis familiaris Loxodonta africana Ovia aries Felis catus	424 453 YTVNILSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE YTVNULSAARMLIH CYKSIANLIERROFE YTVNULSAARMLIH CYKSIANLIERROFE YTVNILSAARMLIH CYKSIANLIERROFE

POLR3C

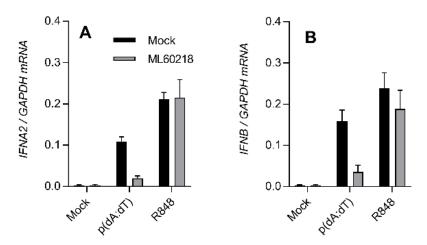
Supplemental Figure 2. Alignments. Alignment across species of the regions surrounding the mutations. Residues marked in red, represent the residue in patients. Residues marked in green represent the residue in the human wildtype allele. Residues marked in yellow are not conserved, but have similar physical properties as the conserved residue at the given position.



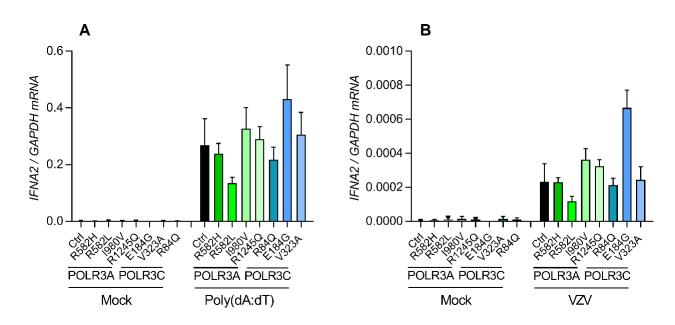
Supplemental Figure 3. Analysis of CADD score versus allele frequency variations in ExAC and VZV patients. (A) *POLR3A*, (B) *POLR3C*. MSC, Mutation Significance Cutoffs.



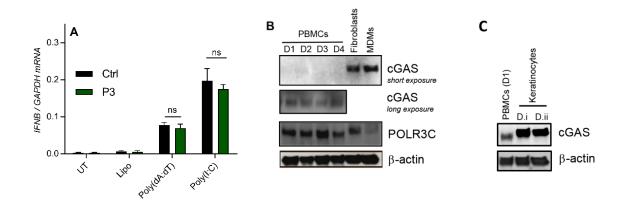
Supplemental Figure 4. Expression of WT and patient POLR3C in bacteria. Bacteria were transduced with plasmids encoding WT POLR3C and the 3 mutant forms from P1, P3, and P4 with a His-tag at the N-terminus. POLR3C expression was induced by addition of 1 mM IPTG treatment. Total lysates were analyzed for expression of POLR3C by Western blot, thus demonstrating that all mutants were expressed (top panel). Lysates were collected and evaluated for protein retention by the His-tag Nickel affinity resin.



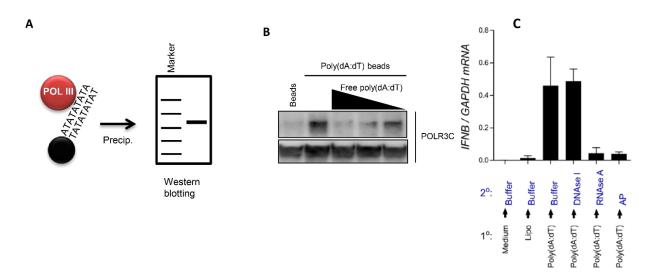
Supplemental Figure 5. POL III-dependent stimulation of IFN expression by poly(dA:dT) in PBMCs. PBMCs from a healthy donor were treated with the Pol III inhibitor ML-60218 (10 μ M) for 16 h prior to transfection of poly(dA:dT) (2 μ g/mL) or treatment with R848 (1 μ g/mL). Total RNA was harvested 6 h after stimulation and levels of (A) *IFNA2* and (B) *IFNB* mRNA were determined by RT-qPCR as shown. Data are shown as means +/- st.dev.



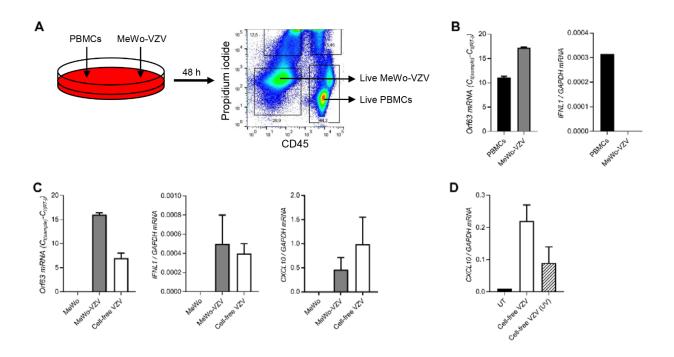
Supplemental Figure 6. Expression of *IFNA2* by PBMCs from donors with other POLR3 mutations. PBMCs from donors with other POLR3 mutations than the ones found in the patients were (A) transfected with poly(dA:dT) ($2 \mu g/mL$), or (B) VZV-infected MeWo cells. Total RNA was harvested 6h after poly(dA:dT) stimulation and 48 h after initiation of co-culture with VZV-MeWo cells. Levels of *IFNA2* mRNA were determined by RT-qPCR as indicated. The levels of cytokine mRNAs were normalized to *GAPDH*, and data are presented as means +/- st.dev. Ctrl, Control. Average of 4 sex- and age-matched controls.



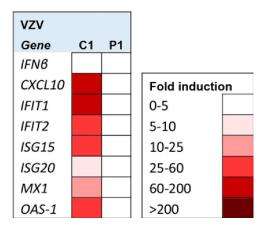
Supplemental Figure 7. Normal response to DNA in fibroblasts from P3. Fibroblasts from P3 and a healthy donor were transfected with (**A**) poly(dA:dT) or poly(I:C) (both 2 μ g/mL). Total RNA was harvested 6 h later and levels of *IFNB* mRNA were determined by RT-qPCR, normalized to *GAPDH*, and presented as means +/- st.dev. ns, p>0.05. (**B**) Total cell lysates from PBMCs (4 donors), skin fibroblasts, and monocyte-derived macrophages were immunoblotted for the levels of cGAS, POLR3C, and β -actin. (**C**) Total cell lysates from PBMCs (1 donor), and keratinocytes (2 donors) were immunoblotted for the levels of cGAS and β -actin.



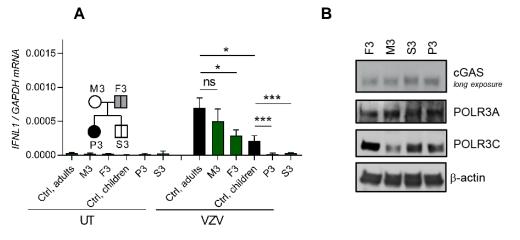
Supplemental Figure 8. Poly(dA:dT) interacts with POL III and leads to generation of immunostimulatory 5'-triphosphorylated RNA. (A) Illustration of the principle steps in precipitation and identification of poly(dA.dT)-interacting proteins. (B) Poly(dA:dT) beads incubated with cytoplasmic extracts of PBMCs from a healthy donor in the presence and absence of increasing levels of free poly(dA:dT) were precipitated and immunoblotted with anti-POLR3C. (C) PBMCs from one healthy donor were treated as indicated under 1°. Total RNA was isolated 14 h later, and the RNA was treated as specified under 2° before transfection into HEK293 cells. Total RNA was harvested 6 h later and levels of *IFNB* mRNA were determined by RT-qPCR and normalized to *GAPDH*. Data are presented as means +/- st.dev.



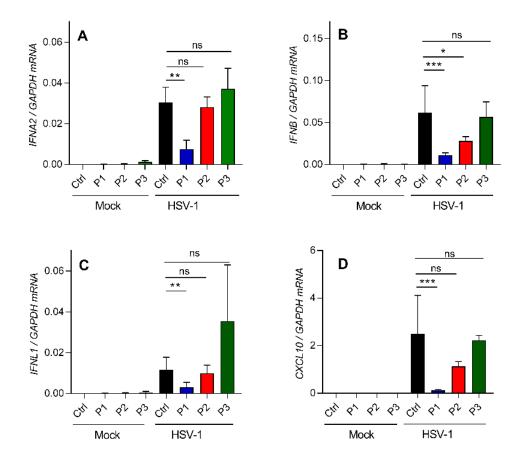
Supplemental Figure 9. Characterization of the PBMC-MeWo coculture system. (**A**, **B**) PBMCs from a healthy donor were co-cultured with MeWo cells (VZV-infected or non-infected) in a 1:1 ratio. The cells two cell populations were separated 48 h later by sorting for CD45 expression, and levels of *Orf63* and *IFNL1* mRNA were determined by RT-qPCR. (**C**) PBMCs were co-cultured with (i) uninfected MeWo cells, (ii) VZV-infected MeWo cells in a 1:1 ratio, or (iii) sonicated and cleared lysates from infected MeWo cells (cell-free VZV). (**D**) In a different experimental set-up PBMCs were co-cultured with cell-free VZV +/- UV-treatment prior to addition to the PBMCs. Total RNA was harvested 12 h after initiation of co-culture/infection, and levels of Orf63, *IFNL1, and CXCL10* mRNA was determined by RT-qPCR. Data are presented as means +/- st.dev.



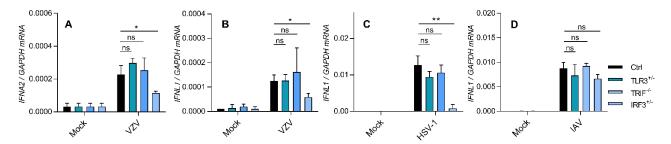
Supplemental Figure 10. Reduced ISG expression in P1 PBMCs after VZV infection. PBMCs from P1 and an age- and gender-matched healthy control were treated with VZV-infected MeWo cells for 48 h. Total RNA was harvested and levels of IFN β and selected ISGs were determined by Fluidigm. The data were normalized to β -actin levels (which were comparable between the two groups) and are shown as heat-maps, with each color point representing means of 3 biological replicates.



Supplemental Figure 11. Characterization of *IFN* λ 1 production in response to VZV infection by PBMCs from the family of P3. (A) PBMCs from P3, family members, and age-matched healthy controls were treated with VZV-infected MeWo cells. Total RNA was harvested 48 h after initiation of co-culture with VZV-MeWo cells and levels of *IFNL1* mRNA were determined by RTqPCR. The levels of the mRNA of interest was normalized to *GAPDH*, and data are presented as means +/- st.dev. Ctrl, Control. Average of 7 and 12 age-matched adult controls in the adult and children groups, respectively. ns, p>0.05; *, 0.01<p<0.05; **, 0.01<p<0.001; ***, 0.001<p<0.0001. (B) Expression levels of cGAS, POLR3A, POLR3C, and β -actin in PBMCs lysates from the family of P3.



Supplemental Figure 12. Expression of IFNs and ISGs by PBMCs from P1-P3 after HSV-1 infection. PBMCs from P1, P2, P3 and healthy controls were infected with HSV-1 (MOI 3). Total RNA was harvested 6 h after infection and levels of (A) *IFNA2*, (B) *IFNB*, (C) *IFNL1*, and (D) *CXCL10* mRNA were determined by RT-qPCR as indicated. The levels of cytokine mRNAs were normalized to *GAPDH*, and data are presented as means +/- st.dev. Ctrl, Control. Average of 4 sexand age-matched controls. ns, p>0.05; *, 0.01 ; **, <math>0.01 ; ***, <math>0.001 .



Supplemental Figure 13. PBMCs from patients with mutations in TLR3 pathway genes have normal response to VZV infection. PBMCs from healthy controls and patients with the indicated mutations in genes of the TLR3 pathway were (**A**, **B**) treated with VZV-infected MeWo cells, (**C**) infected with HSV-1 (MOI 3), or (**D**) IAV (MOI 8). Total RNA was harvested 48 h after initiation of co-culture with VZV-MeWo cells and 6 h after HSV and IAV infections. Levels of *IFNA2* and *IFNL1* mRNA were determined by RT-qPCR as indicated. The levels of the mRNA of interest were normalized to *GAPDH*, and data are presented as means +/- st.dev. Ctrl, Control. Average of 4 agematched controls. ns, p>0.05; *, 0.01<p<0.05; **, 0.01<p<0.001; ***, 0.001<p<0.0001.

Gene symbol	Gene name	HGNC ID	Chromosome Location
ADAR1	Adenosine deaminase	103	1q21.3
AIM2	Absent in melanoma 2	357	1q22
AMFR	Autocrine Motility Factor Receptor	463	16q21
APOBEC3G	apolipoprotein B mRNA editing enzyme catalytic subunit 3G	17357	22q13.1-q13.2
ATF2	Activating transcription factor 2	784	2q32
ATM	Ataxia telangiectasia mutated	795	11q22-q23
BST2	Bone marrow stromal cell antigen 2	1119	19p13.1
BTK	Bruton agammaglobulinemia tyrosine kinase	1133	Xq21.33-q22
CASP1	Caspase 1	1499	11q23
CASP10	Caspase 10	1500	2q33-q34
CASP8	Caspase 8	1509	2q33-q34
<i>CD74</i>	CD74	1697	5q32
CHUK	Conserved helix-loop-helix ubiquitous kinase / Nuclear Factor NF-Kappa-B	1974	10q24-q25
	Inhibitor Kinase Alpha Class II, major histocompatibility		
CIITA	complex, transactivator	7067	16p13
CXCL9	C-X-C motif chemokine ligand 9	4283	4q21
CXCL10	C-X-C motif chemokine ligand 10	3627	4q21
CXCL11	C-X-C motif chemokine ligand 11	6373	4q21.2
DDX41	DEAD (Asp-Glu-Ala-Asp) box polypeptide 41	18674	5q35.3
DDX58	DEAD (Asp-Glu-Ala-Asp) box polypeptide 58 (RIG-I)	23586	9p12
DDX60	DEAD (Asp-Glu-Ala-Asp) box polypeptide 58	25942	4q32.3
DHX9	DEAH-box helicase 9	2750	1q25
DHX36	DEAH-box helicase 36	14410	3q25.2
DHX58	DEAH-box helicase 58 (LGP2)	29517	17q21.2
DOCK2	Dedicator of cytokinesis 2	1794	5q35.1
DOCK8	Dedicator of cytokinesis 8	19191	9p24.3
GBP1	Guanylate binding protein 1	4182	1p22.2
GBP2	Guanylate binding protein 2	4183	1p22.2
GATA2	GATA binding protein 2	4171	3q21
HPSE	Heparanase	5164	4q21.3
IFI6	Interferon alpha inducible protein 6	4054	1p35
IFI16	Interferon, gamma-inducible protein 16	5395	1q22
IFIH1	Interferon induced with helicase C domain 1 (MDA5)	18873	2q24.2
IFIT1	Interferon induced protein with tetratricopeptide repeats 1	5407	10q23.31
IFIT2	Interferon induced protein with tetratricopeptide repeats 2	5409	10q23.31
IFIT3	Interferon induced protein with tetratricopeptide repeats 3	5411	10q24

Supplemental Table 1. Genes involved in IFN production and function

	Interferon induced transmembrane		
IFITM1	Interferon induced transmembrane	5412	11p15.5
	protein 1 Interferon induced transmembrane		
IFITM2	protein 2	5413	11p15.5
	Interferon induced transmembrane		
IFITM3	protein 3	5414	11p15.5
IFNA1	Interferon alpha1	3439	9p22
IFNA2	Interferon alpha2	3440	9p22 9p22
IFNA2 IFNA4	Interferon alpha4	3440	9p22 9p22
IFNA5	Interferon alpha5	3442	9p22 9p22
IFNA6	Interferon alpha6	3443	9p22 9p22
IFNA7	Interferon alpha7	3444	9p22 9p22
IFNA8	Interferon alpha8	3445	9p22 9p22
IFNA10	Interferon alpha10	3446	9p22 9p22
IFNA13	Interferon alpha13	3447	9p22 9p22
IFNA13 IFNA14	Interferon alpha14	3447	9p22 9p22
IFNA16	Interferon alpha16	3449	9p22 9p22
IFNA17	Interferon alpha17	3449	9p22 9p22
IFNA17 IFNA21	Interferon alpha ¹⁷	3451	-
ΙΓΙΝΑΖΙ	*	5452	9p22
IFNAR1	Interferon alpha and beta receptor subunit 1	3454	21q22.11
	Interferon alpha and beta receptor		
IFNAR2	subunit 1	3455	21q22.11
IFNB1	Interferon, beta1	3456	0n21
IFNG		5438	9p21 12q14
IFNGR1	Interferon gamma Interferon gamma receptor 1	5439	6q23-q24
IIIIIONI		5459	0q23-q24
IFNGR2	Interferon gamma receptor 2 (interferon	5440	21q22.1
IFNL1	gamma transducer 1) Interferon lambda 1	18363	10a12 12
IFNLI IFNL2	Interferon lambda 2	18363	19q13.13
IFNL2 IFNL3	Interferon lambda 3	18365	19q13.13
IFNL3 IFNL4	Interferon lambda 4	44480	19q13.13 19q13.12
IFNL4 IFNLR1		18584	19415.12 1p36.11
ΙΓΙΝΔΚΙ	Interferon lambda receptor 1	16364	1050.11
IKBKB	Inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase beta	5960	8p11.2
	Inhibitor of kappa light polypeptide gene		
IKBKE	enhancer in B-cells, kinase epsilon	14552	1q31
	Inhibitor of kappa light polypeptide gene		
IKBKG	enhancer in B-cells, kinase gamma	5961	Xq28
IL10RB	interleukin 10 receptor subunit beta	5965	21q22.11
ILIOND ILIRN	Interleukin 1 receptor antagonist	6000	2q14.2
	Interleukin-1 receptor-associated kinase	0000	2414.2
IRAK1	1	3654	Xq28
	Interleukin-1 receptor-associated kinase		
IRAK2	2	3656	3p25.3
	Interleukin-1 receptor-associated kinase		
IRAK4	4	17967	12q12
IRF1	4 Interferon regulatory factor 1	6116	5q31.1
IRF1 IRF3	Interferon regulatory factor 3	6118	19q13.3-q13.4
IRF5 IRF5	Interferon regulatory factor 5	6210	
IRF3 IRF7	Interferon regulatory factor 7	6122	7q32 11p15.5
IRF7 IRF8	Interferon regulatory factor 7	5358	16q24.1
IMPO	interteron regulatory factor o	5550	10474.1

IRF9	Interform regulatory factor 0	6131	14q11.2
	Interferon regulatory factor 9 Interferon stimulated gene 15, ubiquitin-		-
ISG15	like modifier	4053	1p36.33
	Interferon stimulated gene 20,	61 0 0	
ISG20	exonuclease	6130	15q26
JAK1	Janus kinase 1	6190	1p32.3-p31.3
JAK2	Janus kinase 2	3717	9p24
JAK3	Janus kinase 3	6193	19p13-p12
MB21D1	Mab-21 domain containing 1 (cGAS)	21367	6q13
MAP3K14	Mitogen-activated protein kinase kinase	6853	_
MALJV14	kinase 14	0855	17q21
MAVS	Mitochondrial antiviral signaling protein	29233	20p13
MRE11A	MRE11 meiotic recombination 11	7230	11q21
	homolog A (S. cerevisiae)	1250	11921
MYD88	Myeloid differentiation primary response	7562	3p22
	88		_
MX1	MX dynamin like GTPase 1	7532	21q22.3
MX2	MX dynamin like GTPase 2	7533	21q22.3
NAMPT	Nicotinamide phosphoribosyltransferase	30092	7q22.3
NFKB2	Nuclear factor of kappa light polypeptide	7795	10q24
	gene enhancer in B-cells 2 (p49/p100)		
NFKBIA	Nuclear factor of kappa light polypeptide	7797	14q13
NILLE 11	gene enhancer in B-cells inhibitor, alpha	25727	_
NHEJ1	Nonhomologous end-joining factor 1	25737	2q35
NLRP12 NLRP2	NLR family, pyrin domain containing 12	22938 22948	19q13.42
NLRP2 NLRP3	NLR family, pyrin domain containing 2	16400	19q13.42
NLKP 3	NLR family, pyrin domain containing 3	10400	1q44
NOD2	Nucleotide-binding oligomerization domain containing 2	5331	16q12
OAS1	2'-5'-oligoadenylate synthetase 1	8086	12q24.2
OAS1 OAS2	2'-5'-oligoadenylate synthetase 2	8087	12q24.2 12q24.2
OAS2 OAS3	2'-5'-oligoadenylate synthetase 3	8088	12q24.2 12q24.2
OASL	2'-5'-oligoadenylate synthetase like	8090	12q24.2 12q24.2
PGM3	Phosphoglucomutase 3	8907	6q14.1-q15
PACT	Protein activator of PKR	9438	2q31.2
	Protein kinase activated by double-		
PKR	stranded RNA	2р22-р21	2p22-p21
POLR3A	RNA polymerase III subunit A	30074	10q22.3
POLR3B	RNA polymerase III subunit B	30348	12q23.3
POLR3C	RNA polymerase III subunit C	30076	1q21
POLR3D	RNA polymerase III subunit D	1080	8q21
POLR3E	RNA polymerase III subunit E	30347	16p12.2
POLR3F	RNA polymerase III subunit F	15763	20p11.23
POLR3G	RNA polymerase III subunit G	30075	5q14.3
POLR3H	RNA polymerase III subunit H	30349	22q13.2
POLR3GL	RNA polymerase III subunit G like	28466	1q21.1
POLR3K	RNA polymerase III subunit K	14121	16p13.3
DDVDC	Protein kinase, DNA-activated, catalytic	0412	-
PRKDC	polypeptide	9413	8q11
PYCARD	PYD and CARD domain containing	16608	16p11.2
RELA	RELA proto-oncogene, NF-kB subunit	9955	11q13
RHBDF2	Rhomboid 5 homolog 2 (iRhom2)	20788	17q25.1

In R1threonine kinase 11001210112RIPK3Receptor-interacting serine-threonine kinase 31002114q12RNaseH2Aribonuclease H2 subunit A1851819p13.2RNaseH2Cribonuclease H2 subunit C2411611q13.1RNaseH2Cribonuclease L60411q25RIP4receptor transporter protein 4239923q27.3SAM and HD domain containing350220pter-q12triphosphohydrolase 1113622q32.2-q32.3STAT1Signal transducer and activator of transcription 1, 91kDa113622q32.2-q32.3STAT2Signal transducer and activator of transcription 21136417q21Signal transducer and activator of transcription 3 (acute-phase response1136417q21STAT3transcription 3 (acute-phase response1136617q11.2STAT5ASignal transducer and activator of transcription 5A1136617q11.2STAT5Bsignal transducer and activator of transcription 5B1136717q11.2TANKTRAF family member-associated NFKB activator115622q24-q31TICAM1TOII-like receptor adaptor molecule 1 (TRIF)118484q32TIRAFToII-like receptor adaptor molecule 2118484q32TIRAFToII-like receptor 7156333p21.3TIRAFToII-like receptor 7156333p21.3TRAFToII-like receptor 7156333p21.3TIRAFToII-like receptor 9156333p21.3TIRAF
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TRIM21Tripartite motif containing 211131211p15.5
TRIM22Tripartite motif containing 221636711p15
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TREX1	Three prime repair exonuclease 1	12269	3p21.31
TYK2	Tyrosine kinase 2	12440	19p13.2
UNC93B1	Unc-93 homolog B1 (C. elegans)	13481	11q13.2
VPS45	Vacuolar protein sorting 45 homolog (S. cerevisiae)	14579	1q21.2
XIAP	X-linked inhibitor of apoptosis	592	Xq25
XRCC5	X-ray repair cross complementing 5	12833	2q35
XRCC6	X-ray repair cross complementing 6	4055	22q13.2
ZBP1	Z-DNA binding protein 1	16176	20q13.31
ZDHHC1	Zinc Finger DHHC-Type Containing 1	17916	16q22.1
Lonner	Zine i inger Dinie Type Containing i	17710	10422

	VZV cohort	Control cohort	VZV vs Control
	(21 individuals)	(3724 individuals)	p-value
POLR3A	3	54	0.0020
POLR3C	2	14	0.0046
POLR3A or POLR3C	4	68	0.00036
POLR3A and POLR3C*	2	4	0.0012

Supplemental Table 2. Enrichment of rare POLR3A, POLR3C variations in severe acute VZV infection cohort

Notes:

1. Ethnic origin of the whole-exome/genome-sequenced individuals, from the VZV cohort or from Control cohort, has been assessed by principal component analysis (PCA) using their exome data. The VZV cohort contains 14 Caucasians, 4 North Africans, 1 African, and 2 Americans. The Control cohort was constructed by including individuals from the Human Genetics of Infectious Diseases (HGID) lab no-viral infection cohorts and from the 1000Genome database ⁽¹⁾.

2. The 'Control cohort' consists of 1220 individuals from the no-viral infection cohorts of the HGID lab, and 2504 individuals from the 1000Genome project. Total number of individuals in the cohort of severe acute VZV infection or the Control cohort is indicated under the name of each cohort.

3. Under a POLR3A or POLR3C monogenic model, rare alleles with a MAF<0.0001 in ExAC, and with a CADD score higher than predicted Mutational Significance Cut-off of 95% confidence (13.910 for *POLR3A*, 11.066 for *POLR3C*) were considered in the test. Under a POLR3A and POLR3C digenic model*, rare alleles with a MAF<0.01 in ExAC, and with a CADD score higher than predicted Mutational Significance Cut-off of 95% confidence were considered in the test. 4. Number of individuals from VZV cohort or Control cohort who harbor rare *POLR3A* or *POLR3C* variations is listed in the table.

5. Ethnic-adjusted *POLR3A* and/or *POLR3C* mutation enrichment assessment was performed using logistic regression adjusting on the three first components of the PCA.

Reference:

Genomes Project, C., Auton, A., Brooks, L.D., Durbin, R.M., Garrison, E.P., Kang, H.M., Korbel, J.O., Marchini, J.L., McCarthy, S., McVean, G.A., et al. 2015. A global reference for human genetic variation. *Nature* 526:68-74.

Supplemental Table 3. Clinical data

Patient # (sex, age)	Clinical presentation	Paraclinical findings	MR/EEG findings	Diagnosis and treatment
1 (M, 3 years)	- Fever	- VZV PCR (CSF ¹), $+^2$	- Brain CAT ⁷ , N	Encephalitis
	- Convulsions, day 6	- WBC ³ , 50 cells/µl	- MRI (4 mo), N	Acyclovir 330 mg x3 i.v. for 21 days
	- Loss of consciousness	- Glucose and protein, N ⁴	- EEG, slowed	Day 14: methylprednisolone pulse
	- Hypertonia	. .		therapy for 3 days
	- Anisocoria			
	- Meningism			
2 (M, 5 years)	 Ataxia and nystagmus during chickenpox Chickenpox twice 	- VZV PCR (CSF), ÷ ⁵	MRI: Arnold-Chiari type I with tonsillar herniation	Cerebellitis
3 (M, 5 years)	- diffuse necrotic varicella	VZV PCR (TSM) ⁶ , +		Pneumonitis
- ()-)	- severe pneumonia			ECMO support needed
4 (F, 11 years)	- bitemporofrontal acute headache	- VZV PCR (CSF), +		Encephalitis
	- varicella at 6 years old			

¹ CSF, Cerebrospinal fluid

² +, positive; ÷, negative
³ WBC, White blood cells

⁴N, Normal

⁵ It has been reported that only a minority of patients with VZV cerebellitis harbor detectable levels of viral DNA in the CSF (1). Therefore, since the onset and resolution of the clinical symptoms of cerebellitis coincided with the chickenpox, it was considered highly likely that VZV was the cause of the CNS symptoms.

⁶Tracheal suction material

⁷CAT, Computed Axial Tomography; MRI, *Magnetic resonance imaging*; EEG, Electroencephalography

Rack, A.L. et al. Neurologic varicella complications before routine immunization in Germany. Pediatr. 1. Neurol. 42, 40-48 (2010).

	P1	P2	VZV patient (no POL III mutation)	Normal range
Hematology				
Neutrophils	3.96 x 10 ⁹ cells/L	6.43 x 10 ⁹ cells/L	2.57 x 10 ⁹ cells/L	1.8-8.0 x 10 ⁹ cells/L
Lymphocytes	2.86 x 10 ⁹ cells/L	2.72 x 10 ⁹ cells/L	3.05 x 10 ⁹ cells/L	1.2-6.5 x 10 ⁹ cells/L
Monocytes	0.96 x 10 ⁹ cells/L	0.73 x 10 ⁹ cells/L	0.64 x 10 ⁹ cells/L	0.1-0.8 x 10 ⁹ cells/L
Eosinophils	0.13 x 10 ⁹ cells/L	0.19 x 109 cells/L	0.75 x 10 ⁹ cells/L	0-0.5 x 109 cells/L
Basophils	0.03 x 10 ⁹ cells/L	0.03 x 10 ⁹ cells/L	0.02 x 10 ⁹ cells/L	0.02-0.1x109 cells/L
Flow cytometry				
T cells				
CD3+	1945 cells /µL	1706 cells /µL	1937 cells /µL	700-4200 cells /µL
CD3+CD4+	826 cells /µL	1100 cells /µL	1075 cells /µL	300-2000 cells /µL
CD3+CD8+	889 cells /µL	491 cells /µL	692 cells /µL	300-1800 cells /µL
CD4+/CD8+	0.93	2.24	1.55	0.9 - 2.6
NK cells				
CD16+CD56+CD3÷	191 cells /µL	593 cells /µL	485 cells /µL	90-900 cells /µL
B cells				
CD19+	167 cells /µL	462 cells /µL	492 cells /µL	200-1600 cells /µL
Kappa	53.7% of CD19+	NA	66.9% of CD19+	
Lambda	46.3% of CD19+	NA	33.2% of CD19+	
Kappa/Lambda	1.16	NA	2.02	0.26 - 1.65
CD10	0.5% of CD19+	NA	0.9% of CD19+	
Plasma cells				
CD38++	0.2%	NA	0.1%	
Granulocytes	55.4%	65.1%	65.1%	35-80%

Supplemental Table 4. Immunophenotyping of available patient blood cells

NA, information no available

Patient, mutation	Localization	Comments
P1, POLR3C L11F	Relatively buried in the core of the protein	May alter the local or general folding of POLR3C
P2, POLR3A M307V	At the interface between POLR3A and POLR3C	May perturb the interaction with POLR3C and its partners, POLR3F and POLR3G
P3, POLR3A Q707R	Exposed to the solvent	May perturb the interaction with associated factors
P3, POLR3C R438G	At the interface with POLR3F	May alter the POLR3C-POLR3F interaction
P4, POLR3A R437Q	Exposed to the solvent	May perturb the interaction with associated factors
P4, POLR3C R84Q	Buried in the core of the protein	May alter folding of POLR3C

Supplemental Table 5. Structural characteristics of the POL III mutations

			Poly(dA:dT)				VZV			
Patient	Genotype	IFNA2	IFNB	IFNL1	CXCL10	I	FNA2	IFNL1	CXCL10	Orf63
P1	POLR3C L11F	\mathbb{R}^1	R	R, resc ²	R	N	I	R, resc	R ³	E, resc
P2	POLR3A M307V	R	R	R	R	N	I	Ν	Ν	Ν
P3	POLR3A Q707R,	R	R	R, resc	R	R		R, resc	R	E, resc
	POLR3C R438Q									
P4	POLR3A R437Q	R	R	R	R	R		R	R	Е
	POLR3C R84Q									

Supplemental Table 6. Summary of phenotypes in PBMCs

¹R, reduced compared to controls; N, normal; E, elevated compared to controls ² resc, rescue of phenotype by transduction with wildtype allele ³ VZV-infected PBMCs from P3 displayed reduced expression of a panel of IFN-stimulated genes compared to controls